# Allelopathy as a way to control pests and weeds

Charles Blancher, Thomas Forestier, Aude Geistodt-Kiener, Inès Seka



### **EXEMPLE OF ALLELOPATHY:**

### 2,4-dihydroxy-7-metoxy-1,4-benzoxazine-3,(4H)-one (DIMBOA)

#### What is DIMBOA ?



DIMBOA is a phytoanticipin benzoxanoid. The maximum concentration occurs when the root is 2 weeks old [5].

The benzoxazinoid glucosides are stored in the vacuole as inactive phytoanticipins and the specific glucosidase is deposited in the maize chloroplast .

When pathogen or herbivore attack destroys the integrity of the cell organelles, glucoside (glucose derived molecole) and glucosidase are combined and the toxic aglycone is produced. In maize, the enzymatic release of DIMBOA after wounding is completed within 30 min [6].

#### **Prerequisites:**

"phytoalexins are low molecular weight, antimicrobial compounds that are both synthesized by and accumulated in plants after exposure to microorganisms" "**phytoanticipins** are low molecular weight, antimicrobial compounds that are present in plants before challenge by microorganisms or are produced after infection solely from preexisting

**Benzoxazinoids** are secondary metabolites that are effective in defence and allelopathy. [7]



#### Figure 1: Cross section of a root

- 1- Pith
- 2- Protoxylem,
- 3- Xylem
- 4- Phloem
- 5- Sclerenchyma
- 6- Cortex,
- 7- Epidermis

**ON PHYTOPHAGES** (*Aphids, Ostrinia nubilalis* and others)

DIMBOA in leaves inhibit the development of about 25% of *Ostrinia nubilalis* by the same means [8].

DIMBOA can be transformed in MBOA by some enzymes presents in some insects intestine. DIMBOA acts primarily as a digestive toxin and that the effects of MBOA occurred after digestion. DIMBOA causes a decrease in the approximate digestibility (AD) and the efficiency of conversion of ingested food (ECI) and has no effect on the efficiency of conversion of digested food (ECD). MBOA causes a decrease in both ECI and ECD but has no effect on AD [9].

#### LOCALIZATION

Mostly concentrated in the cortex of all parts of some Maize and others Grasses roots. DIMBOA gets out actively of the cell via the multidrug and toxin extrusion (MATE) transporters and/or passively through the membrane. [10]

#### On Pseudomonas putida

Benzoxazinoids in root exudates of maize attract *P. putida* to the rhizosphere [11]. This rhizobacteria enhance the presence of encoding genes involved in pathways linked to plant growth promoting properties, such as the production of phytohormones. It also enhance defense responses of maize after wounding [8].

#### **Examples of DIMBOA's effects**



### - Induce production of phytohormones involved in plant growth

#### On Fusarium graminearum

DIMBOA strongly affects the expression of Tri6, encoding a major transcriptional regulator of several genes of the trichothecene biosynthesis pathway. DIMBOA also repressed expression of Tri5, encoding trichodiene synthase, the first enzyme in the trichothecene biosynthesis pathway [12].

Trichothecenes are mycotoxins predominantly produced by the phytopathogenic *Fusarium* fungus, and act in plants as virulence factors aiding the spread of the fungus during disease development. Known for their inhibitory effect on eukaryotic protein synthesis, trichothecenes also induce oxidative stress, DNA damage and cell cycle arrest and affect cell membrane integrity and function in eukaryotic cells [13].

#### On Avena sativa

DIMBOA would inhibit the germination and seedling growth of *A. sativa* by inhibiting the gibberellin-induced induction of  $\alpha$ -amylase production [14].

 $\alpha$ -Amylase plays a major role in the degradation of reserve carbohydrate to soluble sugars during germination.

Therefore, the induction of  $\alpha$ -amylase is a requirement for not only seed germination, but also for subsequent seedling growth until photosynthesis is sufficient to support growth [15].

# Limits and ways to improve the use of DIMBOA

#### **Change agricultural practices**

#### Mixture:

Mix Poaceae with other cultures would permit to control pests, bacterias, fungis and weeds.

**Limits**  $\rightarrow$  Maize producing DIMBOA does not have a lower grain yield [16]. But the farmer needs to be able to produce two different crops instead of one, he would need buy or rent other machines or associate himself with an other farmer. It has been proven that in association with some crop, maize does not have negative impact on the other plants,[17] but it has not been proven for all the crop. **Mulch:** 

To reduce seed growth and bacterial development, residues (roots especially) of Paceae varieties could be used as mulch in crops (before seedling).

**Limits**  $\rightarrow$  DIMBOA half life is from 30 to 84 hours depending on the concentration, MBOA's half life is from 4 to 39 days [18].



Association of maize and wheat

Consequently, the mulch can be used for short cultures or as a pesticide.



Schema of the modification of DIMBOA-glc promotor

#### <u>Genetic</u>

One of the main factor that limits the use of DIMBOA is its variations with environmental conditions and plant growth stages.

#### Variety selection

The selection of variety with high concentration of DIMBOA and low environment dependence for DIMBOA synthesis could also be a solution for Maize protection. The locus bx1 is one candidate that can be selected, it explained 12% of DIMBOA concentration variability. [19]

#### Express DIMBOA-glc gen in the adult stage

DIMBOA-glc, the precursor of DIMBOA, has a higher concentration when the plant is young. The biosynthesis stills continue six to ten days after germination and then the concentration drops down due to release, degradation or transformation of the molecule.[20]

The low concentration in adult plant can be an issue for crop protection, especially for aphids attack. Finding a way to reduce the loss of DIMBOA would be a way to improve Poaceae resistance. Maybe by changing the promoter of the gene coding for DIMBOA-glc and choosing a promoter expressed in great quantity in the adult stage.

# Idea of experimentation that could lead to a new use of DIMBOA

#### I - Foreword

DIMBOA repress the virulence of *Fusariums* by inhibiting some mychotoxins biosynthesis *Fusarium graminearum* is an Ascomycète that conserve itself on the ground.

#### II - Context

Rapeseed is a crop in difficulty today, the pesticides are not effective anymore, no solution in proposed yet. The biggest problem is *Sclerotinia sclerotiorum,* is an Ascomycète that conserve itself on the ground.

#### <u>III – Hypothesis 1</u> <u>That we will test</u> here

DIMBOA could potentially have an effect on S. sclerotiorum, inhibiting mycotoxins or by an other way. III' – Hypothesis 2 That we will not test here Maize does not reduce Rapeseed grain yield or less

#### **IV - Protocole to test the Hypothesis**

- Sow 4 Brassica napus L. var. Oléor in 20 pots.
- Sow 4 Maize DIMBOA-producing (*Zea mays L var.B73*) in 10 of them and 4 Maize non-DIMBOA-producing (*Zea mays L var. bxbx*) in the 10 left [21].
- Seperate the two pots from the others but kept in the same controlled environment: 20°C, HR="60%. Grow them one month.
- Inoculate Sclerotinia sclerotiorum in all pots and change the HR to 95% (best S. sclerotiorium germination) [22]. Wait two weeks for S. to make several life cycles.

#### V - Hoped results

The "bxbx" pots are significantly more affected by S. *sclerotiorium* 

#### VII - Field association

 Brassica napus L. var. Oléor with Zea mays L var.B73 in inter-rows.

#### **VI - Hoped Conclusions**

 $\rightarrow$  We can suppose that the yield will be lower for the "bxbx".

than S. sclerotiorium.

→ Maize producing DIMBOA could be used to tackle the *S. sclerotiorum* issue on Rapeseed.

#### **VIII - Perspective**

- Test if *DIMBOA* damages *S. sclerotiorum* sclerotes and spores.
- If yes: sow B73 in June, cut it down in August to plant Rapeseed, harvest it in June etc.

## **References**

[1] Rice, E.L. (1984) Allelopathy. 2nd Edition, Academic Press, New York, 422.

[2] Noor Shah, Adnan & Iqbal, Javaid & Ullah, Abid & Yang, Guozheng & Yousaf, Muhammad & Fahad, Shah & Tanveer, Mohsin & Hassan, Waseem & Tung, Shahbaz & Wang, Leishan & Khan, Aziz & Wu, Yingying. (2016). Allelopathic potential of oil seed crops in production of crops. A Review. Environmental Science and Pollution Research.

[3] Aslam, Farhena & Khaliq, Abdul & Matloob, Amar & Tanveer, Asif & Hussain, Saddam & Zahir, Zahir. (2016. Allelopathy in agro-ecosystems: a critical review of wheat allelopathy-concepts and implications. Chemoecology.

[4] Raymond Reau, Jean-Marie Bodet, Jean-Paul Bordes, Thierry Doré, Sabah Ennaifar, et al. (2005). Effets allélopathiques des Brassicacées via leurs actions sur les agents pathogènes telluriques et les mycorhizes: analyse bibliographique. Partie 1.

[5] Oliveros-Bastidas, Alberto & Macías, Francisco & Molinillo, José. (2012). Variation Endogenus and Exogenous of Allelochemical 2,4-dihydroxy-7-metoxy-1,4-benzoxazin-3,(4H)-one (DIMBOA) in Root Architecture of Maize (Zea mayz). International Journal of Agriculture and Forestry.

[6] Von Rad, Uta & Hüttl, Regina & Lottspeich, Friedrich & Gierl, Alfons & Frey, Monika. (2002). Two glucosyltransferases are involved in detoxification of benzoxazinoids in maize. The Plant journal : for cell and molecular biology.

[7] Monika Frey, Katrin Schullehner, Regina Dick, Andreas Fiesselmann, Alfons Gierl. (2009). Benzoxazinoid biosynthesis, a model for evolution of secondary metabolic pathways in plants. Phytochemistry.

[8] R. Feng, Jon G. Houseman, A.E.R. Downe, J. Atkinson, J.t. Arnason. (1992). Effects of 2,4-dihydroxy-7methoxy-1,4-benzoxazin-3-one (DIMBOA) and 6-methoxybenzoxazolinone (MBOA) on the detoxification processes in the larval midgut of the European corn borer. Pesticide Biochemistry and Physiology.

[9] Jon G. Houseman, F. Campos, N. M. R. Thie, B. J. R. Philogène, J. Atkinson, P. Morand, J. T. Arnason. (1992). Effect of the Maize-Derived Compounds DIMBOA and MBOA on Growth and Digestive Processes of European Corn Borer (Lepidoptera: Pyralidae). Journal of Economic Entomology.

[10] Jitka Frébortová, Ondřej Novák, Ivo Frébort Radek Jorda. (2010) Degradation of cytokinins by maize cytokinin dehydrogenase is mediated by free radicals generated by enzymatic oxidation of natural benzoxazinones. The Plant Journal

[11] Neal AL, Ahmad S, Gordon-Weeks R, Ton J (2012) Benzoxazinoids in Root Exudates of Maize Attract Pseudomonas putida to the Rhizosphere. PLoS ONE 7(4): e35498.

[12] Nasmith CG, Walkowiak S, Wang L, Leung WWY, Gong Y, Johnston A, et al. (2011) Tri6 Is a Global Transcription Regulator in the Phytopathogen Fusarium graminearum. PLoS Pathog 7(9): e1002266

[13] Arunachalam, Chanemougasoundharam & Doohan, Fiona. (2012). Trichothecene toxicity in eukaryotes: Cellular and molecular mechanisms in plants and animals. Toxicology letters.

[14] Pérez, Francisco. (1990). Allelopathic effect of hydroxamic acids from cereals on Avena sativa and A. Fatua. Phytochemistry.

[15]Hisashi Kato-Noguchi (2008). Effects of four benzoxazinoids on gibberellin-induced  $\alpha$ -amylase activity in barley seeds. Journal of Plant Physiology.

[16]Hermann M. Niemeyer. (2009). Hydroxamic Acids Derived from 2-Hydroxy-2H-1,4-Benzoxazin-3(4H)-one: Key Defense Chemicals of Cereals. Journal of Agricultural and Food Chemistry.

[17] Willey, R., & Osiru, D. (1972). Studies on mixtures of maize and beans (Phaseolus vulgaris) with particular reference to plant population. The Journal of Agricultural Science

[18] Francisco A. Macías,\*, Alberto Oliveros-Bastidas,†, David Marín,Diego Castellano,Ana M. Simonet, and, and José M. G. Molinillo. (2004). Degradation Studies on Benzoxazinoids. Soil Degradation Dynamics of 2,4-Dihydroxy-7-methoxy-(2H)-1,4benzoxazin-3(4H)-one (DIMBOA) and Its Degradation Products, Phytotoxic Allelochemicals from Gramineae. Journal of Agricultural and Food Chemistry.

[19] Butrón, A.M., Chen, Y.C., Rottinghaus, G.E., & McMullen, M.D. (2009). Genetic variation at bx1 controls DIMBOA content in maize. Theoretical and Applied Genetics,

[20] Cambier, V; Hance, T; de Hoffmann, E (2000). "Variation of DIMBOA and related compounds content in relation to the age and plant organ in maize". Phytochemistry.

[21] Shivendra V. Sahi, C.E. Anderson, W.S. Chilton. (1995). The corn wound metabolite DIMBOA causes cell death in tobacco and corn. Plant Science.

[22] Annette Penaud. (Mars 2008).Le sclérotinia déroule son cycle. Perspectives agricoles n°343.